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### Design, synthesis, docking studies, and antibiotic evaluation (*in vitro*) of some novel triphenylamine chalcones and their analogues



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#### ABSTRACT

**Background:** Antibiotic resistance has risen as a result of a variety of conditions, prompting researchers to look for new compounds that can combat multidrug-resistant organisms. Over the last two decades, chalcones have been proved to be attractive moieties in drug discovery. Various substituted acetophenones, propiophenones, and 4-(diphenylamino) benzaldehyde were combined, using the aldol condensation reaction to obtain eight novel triphenylamine chalcones. The compound's antimicrobial properties were investigated (*in vitro*). With the nonmutant X-ray target receptor (PDB: 5VBU), molecular docking experiments were also carried out to analyze the most favorable conformation and find the orientation that maximizes interaction and minimizes energy.

**Results:** Eight novel triphenylamine chalcones were successfully synthesized and recrystallized using ethanol; the percentage yield of the compounds was between 30% and 92%. The activity against different pathogens revealed that all synthesized compounds showed marked antimicrobial activity against the tested microorganisms. Compound **1b** showed the highest zone of inhibition (ZOI) against *Aspergillus niger*, measuring 30 mm. The minimum inhibitory concentration (MIC) results revealed that compounds **1a–1d**, and **2d** had the lowest MIC and inhibited *A. niger* growth at 12.5 g/ml. All the synthesized compounds showed a minimum bactericidal/fungicidal concentration (MBC/MFC) effect against *Escherichia coli*, *Pseudomonas aeruginosa*, *Bacillus subtilis*, *Candida albicans*, and *A. niger* at 50 µg/ml. The docking studies of the synthesized chalcones with the binding site of the target receptor reveal that the binding affinity of the synthesized chalcones was in the range of –11.2 to –9.4 kcal/mol, and showed the highest binding score compared to that of the standard drugs (fluconazole and ciproflaxacin), with docking scores of –7.9 and –7.3 kcal/mol, respectively.

**Conclusion:** The investigation reveals that compound **1b** showed the highest ZOI of 30 mm, least MIC, and MBC/MFC of 12.5 and 50 µg/ml, respectively, against *A. niger*, therefore displaying better antifungal potential as compared to the rest of the compounds. The outcome of the docking analysis revealed that compound **2a** showed a better binding affinity of –11.2 kcal/mol, which is higher than the remaining compounds and the control drugs (fluconazole and ciproflaxacin). Therefore, compounds **1b** and **2a**, which showed better antifungal and highest binding affinity, could be potential candidates in drug design.

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## 1. Background

The chemistry of chalcones has generated a bustle of research around the globe. The synthesis, chemical reactions, and biological applications of these compounds have captivated the attention of researchers (Kumara et al., 2017; Singh et al., 2014). Chalcones are characterized by a conjugated double bond on both benzene rings and a totally delocalized  $\pi$ -electron system (Maidur and Patil, 2018), which is dependent on the presence of other auxochromes.  $\beta$ -phenyl- $\alpha$ -benzoyl ethylene is another name for chalcones (Ugwu et al., 2015). Chalcones are abundantly found in spices, tea, fruits, and vegetables as one of the principal classes of natural products (Kumar et al., 2021). Claisen–Schmidt condensation of acetophenones with benzaldehydes is the most prevalent and efficient methods for producing chalcones. The yields are typically excellent, and reaction takes place at a lower temperature. The duration for reaction varies from 3 to 20 hours, depending on the substitution pattern and the solvent/base system utilized (Adnan et al., 2020; Bukhari et al., 2013).

Chalcones have proven to be biologically active; some substituted derivatives, including heterocyclic analogs, have been found to exhibit strong biological characteristics that have been shown to inhibit micro-organism development (Frazier, 2020). Some chalcone derivatives have proven to be poisonous to mammals (Gomes et al., 2009) and insects (Singh et al., 2019), as well as inhibit enzymes (Agilandeshwari et al., 2016) and herbaceous plants (Mahapatra et al., 2019). They possess anti-inflammatory (Rashid et al., 2019; Ugwu et al., 2015), antifungal (Singh et al., 2019), antioxidant (Kumar et al., 2020; Singh et al., 2019; Vagish et al., 2021), anti-ulcer (Ugwu et al., 2015), antimalarial (Reeta et al., 2019), antituberculosis (Matos et al., 2014), antitumor (Ugwu et al., 2015), anti-helminthic (Matos et al., 2014), antispasmodic (Ugwu et al., 2015), antibacterial (Singh et al., 2019), and antineoplastic activities (Ugwu et al., 2015). These are just a few of the many biological activities linked to chalcones. Antimalarial action has been found for quino-line-based chalcones (Karaman et al., 2010; Martelli et al., 2019; Tomar et al., 2010; Ugwu et al., 2015).

Due to the continued resistance and resurgence of pathogens against available antibiotics, the constant search for potentially novel drug candidates has emerged. Simultaneously, researchers developed the concepts of protein purification and crystallography, allowing researchers to learn more about the protein and ligand interactions. Today, computational methodologies are pervading many facets of

drug development (Bajorath, 2002; Jorgensen, 2004; Walters et al., 1998). Molecular docking investigations have become important tools in the search for new drugs candidates (Langer and Hoffmann, 2001). This technique represents the atomic level interface with target proteins, allowing us to define small molecule behavior in target protein binding sites (McConkey et al., 2002). The process also determines the ligand structure, location, and orientation within the binding site, as well as the binding affinity (Kitchen et al., 2004; Prabhudeva et al., 2019). To our knowledge, there is no report on synthesis, antibiotic evaluation, and molecular docking studies of triphenylamine chalcones. Hence, the present research intended to design, synthesize, screen antimicrobial potential, and study the molecular docking of novel triphenylamine chalcones.

## 2. Methods

Reagents and solvents used in this research were obtained from Sigma-Aldrich (Germany). These were used as obtained without further purification.

### 2.1. Synthesis of the novel triphenylamine chalcones analogs (1a–d, 2a–d)

To a 100 ml round-bottom flask, equipped with a magnetic stirrer, dissolved in 25 ml ethanol, equimolar quantities of 4-(diphenylamino)benzaldehyde (0.4 g, 1.5 mmol) and substituted acetophenones/propionophenone (0.4 g, 2 mmol) were transferred and stirred for 30 minutes. A solution of sodium hydroxide (10 ml, 10%) was added dropwise while stirring. The temperature for the reaction was maintained between 20°C and 24°C using cold water bath for 4–5 hours. The progress and completion of the reaction was monitored using the thin-layer chromatographic technique. On completion, the reaction mixture was placed in a refrigerator for 10 hours. Formation of a precipitate was observed, which was filtered, washed several times with water (100 ml), dried in air, and purified by recrystallization from ethanol (30 ml) to obtain a triphenylamine chalcone (1a–d, 2a–d) (Table 1) (Hongtian et al., 2019; Schemes 1 and 2).

### 2.2. Chemistry

The synthesis of the target novel triphenylamine chalcones was achieved via the conventional Claisen–Schmidt condensation reaction, where different equimolar substituted acetophenones/propionophenones and 4-(diphenylamino)benzaldehyde in the presence of NaOH (10%) were stirred. This reaction was

originated by the abstraction of protons from the alpha-carbon of the acetophenones/propionophenones to generate the resonance stabilized enolate ion by the base. The second stage was the nucleophilic attack on the electron-deficient carbonyl carbon of 4-(diphenylamino)benzaldehyde, which results in the formation of a new C–C bond. This connects the alpha-carbon of the acetophenones/propionophenones to the aldehydic or carbonyl carbon of 4-(diphenylamino)benzaldehyde, which forms an intermediate. The reaction was completed by protonation and deprotonation by the hydroxyl ion of the base to form the targeted  $\alpha,\beta$ -unsaturated triphenylamine chalcone.

### 2.3. Antimicrobial evaluation

The synthesized triphenylamine chalcones were screened for antimicrobial properties, against some selected clinical isolates, which included *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Bacillus subtilis*, *Candida albicans*, *Salmonella typhi*, *Aspergillus niger*, *Escherichia coli*, and methicillin resistant *S. aureus* (MRSA).

#### 2.3.1. Zone of inhibition (ZOI)

The ZOI of the synthesized triphenylamine chalcones was carried out according to the procedure as described by Karou et al. (2006).

#### 2.3.2. Minimum inhibitory concentration (MIC)

The MIC of the synthesized triphenylamine chalcones was determined by using the broth dilution method as reported by Bruton et al. (2007).

#### 2.3.3. Minimum bactericidal/fungicidal concentration (MBC/MFC)

The MBC/MFC of the synthesized triphenylamine chalcones were determined according to the procedure, as described by the Clinical and Laboratory Standard Institute (CLSI, 2015).

### 2.4. Molecular docking studies

Molecular docking analysis for eight ligands (novel triphenylamine chalcones analogs) and controls (fluconazole and ciproflaxacin) was carried out to examine the most favorable interaction and identify the orientation which maximizes interactions and minimizes energy with the target receptor (PDB: 5VBU) (www.rcsb.org) (Fig. 1).

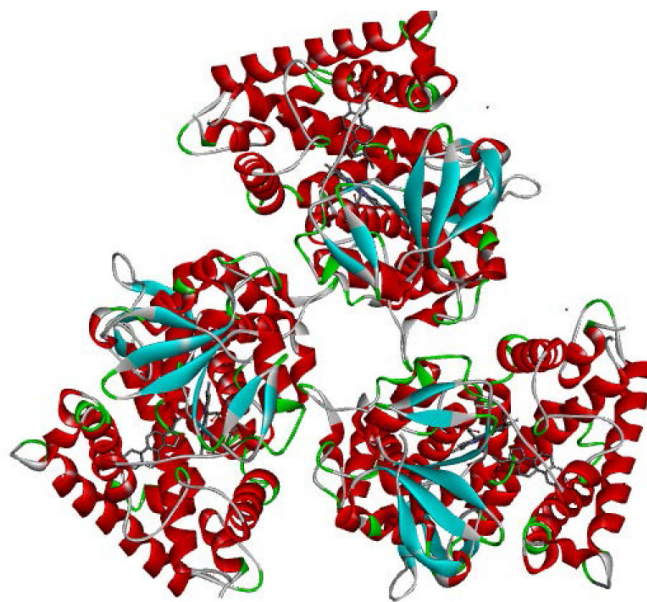
#### 2.4.1. Preparation of the receptor

The three-dimensional structure of the target receptor (Fig. 1) retrieved from the PDB was prepared by

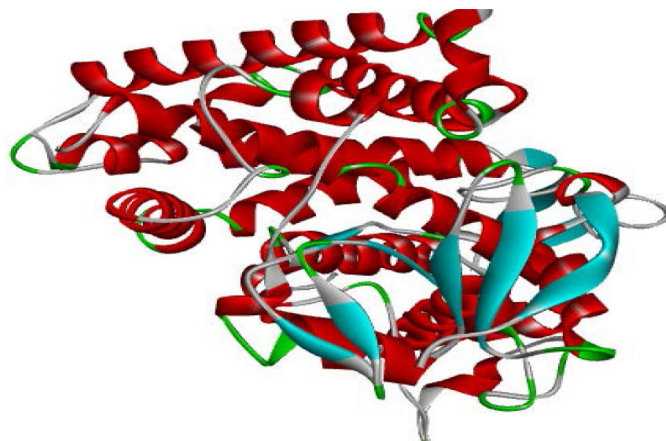
removing the water molecules and other heteroatoms, before minimization for the docking study. Discovery Studio Visualizer software v.21.1.0.20298 was used for the preparation. The treated target receptor (Fig. 2) was saved in pdb file format and transferred to Pyrx software for docking.

#### 2.4.2. Preparation of the ligand

All the eight ligands (triphenylamine chalcones analogs) and controls (fluconazole and ciproflaxacin) studied were designed and synthesized as mentioned earlier. ChemDraw ultra 8.0 software was used to



**Figure 1.** Crystal structure of human cytochrome P450 21A2 hydroxyprogesterone complex (PDB: 5VBU) showing the attached ligands and chains A, B, and C (Chumxue et al., 2017).



**Figure 2.** 3D structure of the optimized chain A of human cytochrome P450 21A2 hydroxyprogesterone complex.

generate the two-dimensional structures of the synthesized chalcones (Li et al., 2004). Spartan software was used to convert the 2D structures to 3D. Geometrical optimization using the AM1 semi-empirical method was carried out on all the compounds using the Spartan software and saved as pdb files.

Polar hydrogens were added before computing the Gasteiger charges.

### 2.4.3. Molecular docking

PyRx virtual screening software was used to study the ligand–receptor interactions between the

**Table 1.** Synthesized triphenylamine chalcones and their percentage yields (%).

| Compounds code | R | Triphenylamine chalcone | Yield |
|----------------|---|-------------------------|-------|
| 1a             |   |                         | 70    |
| 1b             |   |                         | 92    |
| 1c             |   |                         | 57    |
| 1d             |   |                         | 60    |
| 2a             |   |                         | 55    |
| 2b             |   |                         | 30    |
| 2c             |   |                         | 52    |
| 2d             |   |                         | 54    |



target receptor (PDB: 5VBU) and the eight synthesized ligands (novel triphenylamine chalcones analogs) and controls (fluconazole and ciproflaxacin), while the Discovery Studio Visualizer software was employed in visualizing and analyzing the docking results.

### 3. Results

#### 3.1. Results of antimicrobial studies

#### 3.2. Results of molecular studies

### 4. Discussion

#### 4.1. Antimicrobial studies

All the synthesized compounds were shown to possess remarkable activities against the tested microbes by showing significant ZOI (Table 2) relative to that of the standard drugs used. Compound **1b** showed a prominent ZOI of 30 mm against *A. niger*, while compounds **1a**, **2d**, **1c**, **2b**, and **2c** showed ZOI of 25, 23, 22, and 21 mm also, against *A. niger*. Compounds

**1c**, **1d**, **2a–2c** each showed ZOI of 20 mm against *B. subtilis*, *P. aeruginosa*, *C. albicans*, and *A. niger*, while compounds **1a**, **1b**, and **2a** showed the lowest ZOI of 13 mm each against *C. albicans*, *S. aureus*, and *E. coli*. Thus, compounds **1a–1d** showed ZOI (25, 30, and 23 mm) greater than the standard drugs (fluconazole and ciproflaxacin) against *A. niger*.

The results of the MIC (Table 3) showed that compounds **1a–1d**, and **2d** possess the least MIC and inhibit the growth of *A. niger* at 12.5 µg/ml, while compounds **1b–1d**, **2a–2d** inhibit the growth of *C. albicans*, *B. subtilis*, MRSA, *E. coli*, *P. aeruginosa*, and *A. niger* at 25 µg/ml. At 50 µg/ml, the growth of *P. aeruginosa*, *B. subtilis*, *C. albicans*, MRSA, *S. typhi*, *S. aureus*, and *E. coli* was inhibited by compounds **1a–1d**, **2a–2d**. Compounds **1a–1d**, **2a–2d** inhibit the growth of *E. coli*, *P. aeruginosa*, *S. typhi*, *S. aureus*, *B. subtilis*, and MRSA at 100 µg/ml.

The results of MBC/MFC (Table 4) showed that compounds **1a–1d**, **2a–2d** completely killed *E. coli*, *P. aeruginosa*, *B. subtilis*, *C. albicans*, and *A. niger* at 50 µg/

**Table 2.** ZOI (mm) for the synthesized triphenylamine chalcones and standard controls against test microbes.

| Test organisms       | Triphenylamine chalcones |    |    |    |    |    |    |    | Cip | Flu |
|----------------------|--------------------------|----|----|----|----|----|----|----|-----|-----|
|                      | 1a                       | 1b | 1c | 1d | 2a | 2b | 2c | 2d |     |     |
| <i>E. coli</i>       | 14                       | 16 | 15 | 20 | 13 | 18 | 19 | 16 | 30  | -   |
| <i>P. aeruginosa</i> | 15                       | 18 | 18 | 18 | 16 | 18 | 20 | 19 | 40  | -   |
| <i>S. typhi</i>      | 16                       | 14 | 17 | 16 | 16 | 13 | 15 | 18 | 26  | -   |
| <i>S. aureus</i>     | 13                       | 15 | 14 | 18 | 13 | 13 | 15 | 15 | 28  | -   |
| <i>B. subtilis</i>   | 16                       | 17 | 20 | 17 | 18 | 20 | 20 | 18 | 25  | -   |
| MRSA                 | 14                       | 16 | 19 | 15 | 16 | 17 | 16 | 19 | 30  | -   |
| <i>C. albicans</i>   | 17                       | 13 | 20 | 13 | 16 | 20 | 20 | 18 | -   | 20  |
| <i>A. niger</i>      | 25                       | 30 | 23 | 20 | 20 | 22 | 21 | 23 | -   | 22  |

-: Not determined, Cip: ciproflaxacin, Flu: Fluconazole.

**Table 3.** MICs (µg/ml) for the synthesized triphenylamine chalcones against the test microbes.

| Test organisms       | Triphenylamine chalcones |      |      |      |     |     |     |      |
|----------------------|--------------------------|------|------|------|-----|-----|-----|------|
|                      | 1a                       | 1b   | 1c   | 1d   | 2a  | 2b  | 2c  | 2d   |
| <i>E. coli</i>       | 100                      | 100  | 100  | 25   | 100 | 50  | 25  | 100  |
| <i>P. aeruginosa</i> | 100                      | 50   | 50   | 50   | 100 | 25  | 25  | 25   |
| <i>S. typhi</i>      | 100                      | 100  | 50   | 100  | 100 | 100 | 100 | 50   |
| <i>S. aureus</i>     | 100                      | 100  | 100  | 50   | 100 | 100 | 100 | 100  |
| <i>B. subtilis</i>   | 100                      | 50   | 25   | 25   | 25  | 25  | 25  | 50   |
| MRSA                 | 100                      | 50   | 25   | 50   | 100 | 50  | 50  | 25   |
| <i>C. albicans</i>   | 50                       | 25   | 25   | 50   | 50  | 25  | 25  | 50   |
| <i>A. niger</i>      | 12.5                     | 12.5 | 12.5 | 12.5 | 25  | 25  | 25  | 12.5 |

**Table 4.** MBC/MFCs ( $\mu\text{g/ml}$ ) of the synthesized triphenylamine chalcones against the test microbes.

| Test organisms       | Triphenylamine chalcones |     |     |     |     |     |     |     |
|----------------------|--------------------------|-----|-----|-----|-----|-----|-----|-----|
|                      | 1a                       | 1b  | 1c  | 1d  | 2a  | 2b  | 2c  | 2d  |
| <i>E. coli</i>       | -                        | -   | -   | 50  | -   | 100 | 100 | -   |
| <i>P. aeruginosa</i> | -                        | 100 | 100 | 100 | -   | 100 | 50  | 100 |
| <i>S. typhi</i>      | -                        | -   | 100 | -   | -   | -   | -   | 100 |
| <i>S. aureus</i>     | -                        | -   | -   | 100 | -   | -   | -   | -   |
| <i>B. subtilis</i>   | -                        | 100 | 100 | 50  | 100 | 50  | 50  | 100 |
| MRSA                 | -                        | -   | 100 | 100 | -   | 100 | -   | 100 |
| <i>C. albicans</i>   | 100                      | 100 | 50  | 100 | 100 | 50  | 50  | 100 |
| <i>A. niger</i>      | 50                       | 50  | 50  | 50  | 50  | 50  | 50  | 50  |

–: No MBC/MFC.

ml, while a concentration of 100  $\mu\text{g/ml}$  was required by compounds **1a–1d**, **2a–2d** to completely kill *E. coli*, *P. aeruginosa*, *S. typhi*, MRSA, *S. aureus*, *B. subtilis*, and *C. albicans*.

#### 4.2. Molecular docking studies

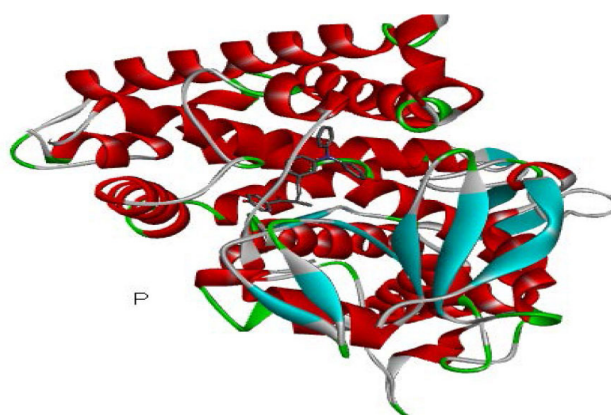
Each ligand/chalcones was successively docked to the binding site of the receptor (PDB: 5VBU) in order to understand the mode of interaction of the triphenylamine chalcones with the target receptor. The analysis of the docking investigations provided us with an insight into the interactional relationship of the novel triphenylamine chalcone analogs and human cytochrome P450 21A2 hydroxyprogesterone complex.

Table 5 showed the results of the binding affinity of the synthesized triphenylamine chalcones/ligands, which ranges between  $-11.2$  and  $-9.4$  kcal/mol. Compound **2a** showed the highest docking score of  $-11.2$  kcal/mol, followed by compounds/ligands **1b** and **2b** which showed the binding energy of  $-10.7$  kcal/mol, while compounds **1c**, **2c**, **2d**, and **1a** showed the docking scores of  $-10.4$ ,  $-10.3$ ,  $-10.3$ , and  $-10.1$ , respectively. Compound/ligand **1d** showed the least binding score of  $-9.4$ . The eight ligands (novel triphenylamine chalcones analogs) showed the highest binding score compared to that of the standard ligand/compounds (fluconazole and ciproflaxacin), which showed the docking scores of  $-7.9$  and  $-7.3$  kcal/mol, respectively.

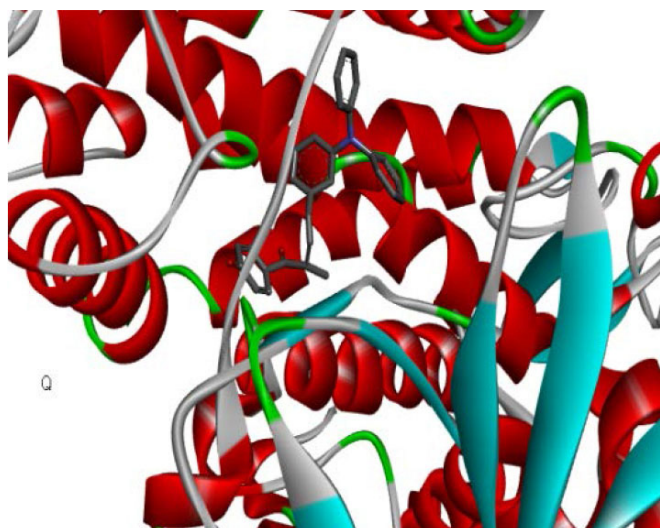
Ligand/triphenylamine chalcone **2a** showed the highest binding affinity ( $-11.2$  kcal/mol) compared to other compounds. Discovery Studio Visualizer software was used to view and analyze the docking investigations. The interaction of the ligand/triphenylamine chalcone (**2a**) with the target receptor (PDB: 5VBU) is shown in Figures 3–5. The interaction

**Table 5.** Binding energy results of the synthesized triphenylamine chalcones with the target receptor (PDB: 5VBU).

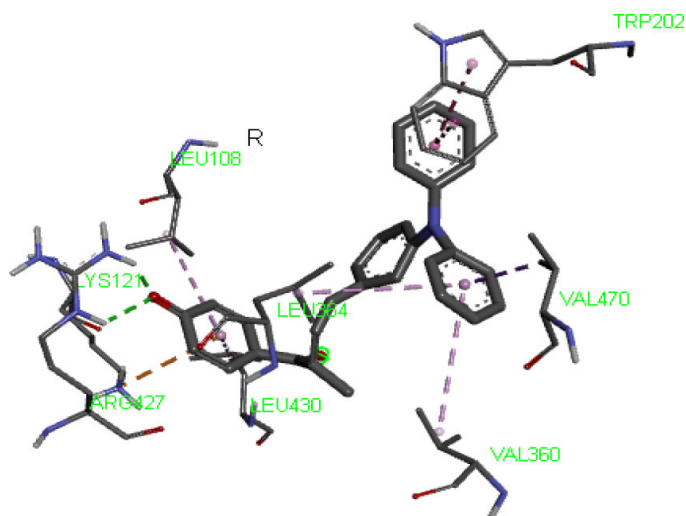
| Triphenylamine chalcones | Binding energy (kcal/mol) |
|--------------------------|---------------------------|
| 1a                       | $-10.1$                   |
| 1b                       | $-10.7$                   |
| 1c                       | $-10.4$                   |
| 1d                       | $-9.4$                    |
| 2a                       | $-11.2$                   |
| 2b                       | $-10.7$                   |
| 2c                       | $-10.3$                   |
| 2d                       | $-10.3$                   |
| Fluconazole              | $-7.9$                    |
| Ciproflaxacin            | $-7.3$                    |

**Figure 3.** 2a-receptor complex in 3D.

was observed with one hydrogen, one electrostatic attraction, and six hydrophobic interactions (2.5449, 4.0569, 3.73562, 4.34337, 5.2863, 5.48907, 5.44454, and 5.35682 Å with ARG427, LYS121, VAL470, TRP202, VAL360, LEU364, LEU108, and LEU430), as

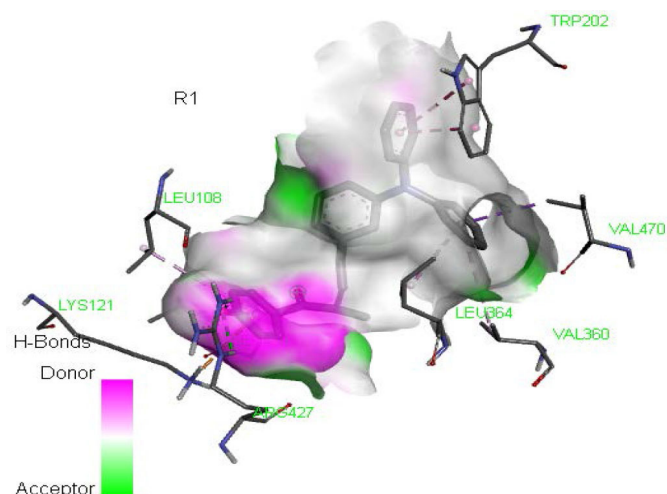


**Figure 4.** Expanded 2a-receptor complex in 3D.

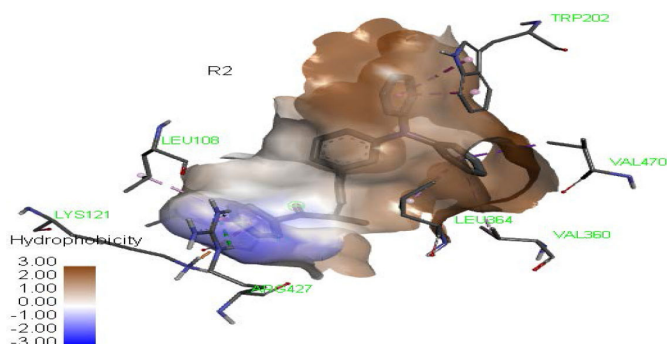


**Figure 5.** 2a-receptor complex showing different interactions in 3D.

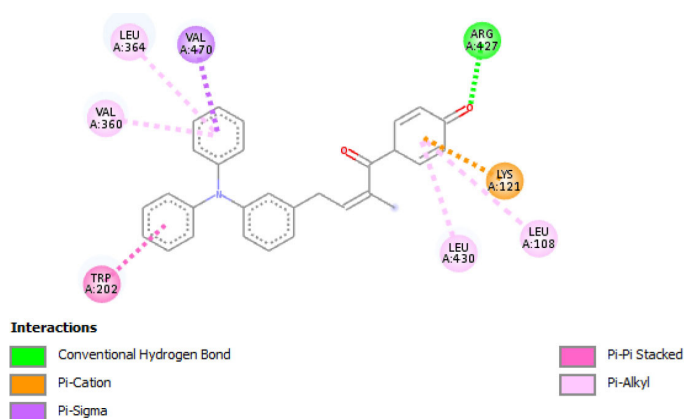
shown in [Figures 6 and 7](#). The two-dimensional structural form of the ligand–receptor complex is shown in [Figure 8](#). One hydrogen bond interaction with ARG427 was formed due to the presence of OH the triphenylamine chalcone/chalcone (**2a**). The electrostatic interaction of the ligand/triphenylamine chalcone was detected with LYS121. Also, the hydrophobic interactions were noticed with VAL470, TRP202, VAL360, LEU364, LEU108, and LEU430 of the target receptor. The areas which represent both hydrogen bond and hydrophobic interactions between the target receptor and the ligand/triphenylamine chalcone (**2a**) fit perfectly into the binding site of the receptor (PDB: 5VBU) ([Table 6](#)).



**Figure 6.** 2a-receptor complex in 3D showing hydrogen bonding.

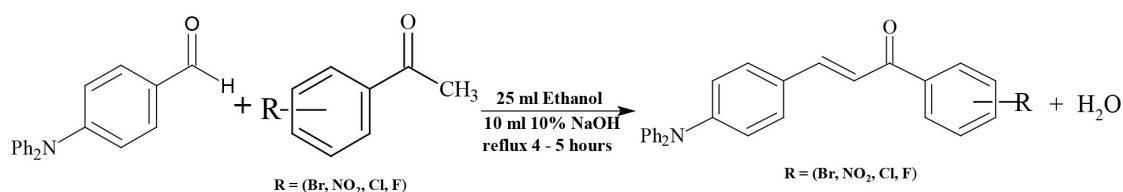


**Figure 7.** 2a-receptor complex in 3D showing hydrophobic interactions.

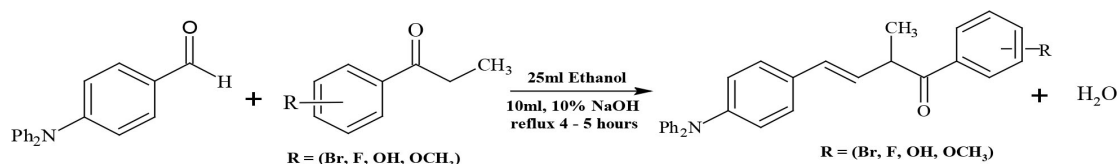


**Figure 8.** 2a-receptor complex in 2D showing types of bonds and interactions.

chalcone (**2a**) fit perfectly into the binding site of the receptor (PDB: 5VBU) ([Table 6](#)).



**Scheme 1.** Synthesis of triphenylamine chalcone from substituted acetophenones (1a–d).



**Scheme 2.** Synthesis of triphenylamine chalcones from substituted propiophenones (2a–d).

**Table 6.** Amino acid interaction, bond distance, and type of ligand–receptor interaction.

| Ligand/compound | Protein interaction | Bond distance (Å) | Type of bond interaction |
|-----------------|---------------------|-------------------|--------------------------|
| <b>1a</b>       | TRP202              | 4.93594           | Hydrophobic              |
|                 | VAL101              | 4.97629           | Hydrophobic              |
|                 | VAL198              | 5.11692           | Hydrophobic              |
|                 | VAL360              | 5.2578            | Hydrophobic              |
|                 | CYS429              | 5.37315           | Hydrophobic              |
|                 | LEU430              | 5.13073           | Hydrophobic              |
|                 | LEU108              | 5.23057           | Hydrophobic              |
|                 | LEU110              | 5.32372           | Hydrophobic              |
| <b>1b</b>       | ARG92               | 2.22588           | Hydrogen bond            |
|                 | ARG427              | 1.83408           | Hydrogen bond            |
|                 | LEU430              | 3.21564           | Hydrogen bond            |
|                 | VAL470              | 3.86767           | Hydrophobic              |
|                 | TRP202              | 3.83466           | Hydrophobic              |
|                 | TRP202              | 4.53881           | Hydrophobic              |
|                 | LEU108-SER109       | 5.28813           | Hydrophobic              |
|                 | VAL428-CYS429       | 5.18984           | Hydrophobic              |
|                 | LEU110              | 5.48518           | Hydrophobic              |
|                 | VAL360              | 5.10827           | Hydrophobic              |
|                 | LEU108              | 5.29357           | Hydrophobic              |
|                 | LEU430              | 5.0342            | Hydrophobic              |
| <b>1c</b>       | VAL198              | 4.6017            | Hydrophobic              |
|                 | LEU227              | 5.18326           | Hydrophobic              |
|                 | CYS429              | 4.28371           | Hydrophobic              |
|                 | VAL360              | 4.8503            | Hydrophobic              |
|                 | ILE471              | 5.47831           | Hydrophobic              |
|                 | VAL101              | 5.41637           | Hydrophobic              |
|                 | ILE291              | 5.32346           | Hydrophobic              |
|                 | TRP202              | 4.24247           | Hydrophobic              |
| <b>1d</b>       | LEU64               | 3.17822           | Halogen                  |
|                 | PRO93               | 3.24979           | Hydrophobic              |
|                 | PRO95               | 3.67448           | Hydrophobic              |
|                 | VAL69               | 4.27875           | Hydrophobic              |
|                 | LEU40               | 4.96375           | Hydrophobic              |
|                 | VAL212              | 5.16423           | Hydrophobic              |
|                 | LEU362              | 4.85287           | Hydrophobic              |
|                 | ILE386              | 4.62883           | Hydrophobic              |
|                 | PRO365              | 5.40796           | Hydrophobic              |
|                 | VAL384              | 5.17048           | Hydrophobic              |
|                 | TYR98               | 4.2068            | Hydrophobic              |

Continued



|                      |        |         |                       |
|----------------------|--------|---------|-----------------------|
| <b>2a</b>            | ARG427 | 2.5449  | Hydrogen bond         |
|                      | LYS121 | 4.0569  | Electrostatic         |
|                      | VAL470 | 3.73562 | Hydrophobic           |
|                      | TRP202 | 4.34337 | Hydrophobic           |
|                      | VAL360 | 5.2863  | Hydrophobic           |
|                      | LEU364 | 5.48907 | Hydrophobic           |
|                      | LEU108 | 5.44454 | Hydrophobic           |
| <b>2b</b>            | LEU430 | 5.35682 | Hydrophobic           |
|                      | LEU64  | 5.08451 | Hydrophobic           |
|                      | ILE386 | 4.65396 | Hydrophobic           |
|                      | PRO95  | 5.23623 | Hydrophobic           |
|                      | PRO365 | 4.89571 | Hydrophobic           |
|                      | LEU40  | 4.99849 | Hydrophobic           |
| <b>2c</b>            | ILE209 | 5.41451 | Hydrophobic           |
|                      | ILE291 | 3.51114 | Hydrophobic           |
|                      | VAL101 | 4.15088 | Hydrophobic           |
|                      | VAL198 | 4.80089 | Hydrophobic           |
|                      | VAL360 | 5.35382 | Hydrophobic           |
|                      | LEU108 | 5.29535 | Hydrophobic           |
|                      | CYS429 | 5.47704 | Hydrophobic           |
| <b>2d</b>            | LEU430 | 4.95443 | Hydrophobic           |
|                      | ARG234 | 2.91932 | Hydrogen bond-halogen |
|                      | VAL360 | 3.43541 | Hydrophobic           |
|                      | CYS429 | 4.3062  | Hydrophobic           |
|                      | LEU199 | 5.46418 | Hydrophobic           |
|                      | ILE471 | 5.47336 | Hydrophobic           |
|                      | VAL101 | 4.66462 | Hydrophobic           |
| <b>Fluconazole</b>   | ILE291 | 5.09895 | Hydrophobic           |
|                      | ASP288 | 2.18916 | Hydrogen bond         |
|                      | VAL470 | 2.64556 | Hydrogen bond         |
|                      | TRP202 | 4.34172 | Hydrophobic           |
| <b>Ciproflaxacin</b> | LEU110 | 3.88479 | Hydrophobic           |
|                      | SER469 | 2.57816 | Hydrogen bond         |
|                      | LEU40  | 2.33333 | Hydrogen bond         |
|                      | GLN42  | 2.66041 | Hydrogen bond         |
|                      | LEU40  | 4.92581 | Hydrophobic           |
|                      | LEU41  | 5.17388 | Hydrophobic           |

## 5. Conclusion

The synthesis of eight novel triphenylamine chalcones was successively carried out via the conventional Claisen–Schmidt condensation reactions. The results of the antimicrobial screening of the synthesized chalcones as indicated by the ZOI showed that all the synthesized compounds possess remarkable activities against the tested microbes, by showing a significant ZOIs relative to that of the standard drugs used. Compound **1b** showed the highest ZOI of 30 mm, least MIC, and MBC/MFC of 12.5 and 50 µg/ml, respectively, against *A. niger*. The outcome of the docking studies revealed that compound **2a** showed marked docking score with binding affinity of –11.2 kcal/mol, which is higher relative to other compounds and the standard controls (fluconazole and ciproflaxacin). Therefore, compounds **1b** and **2a**, which showed better antifungal

and highest binding affinity, could be potential candidates in drug design.

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